

The Synthesis and Biochemical Evaluation of New A₁ Selective Adenosine Receptor Agonists Containing 6-Hydrazinopurine Moieties

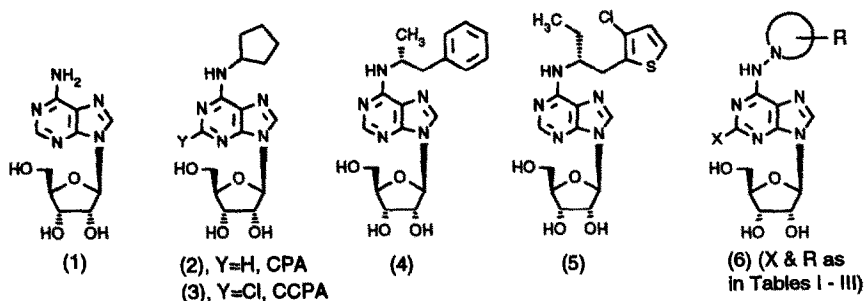
Lars J.S. Knutsen*, Jesper Lau, Malcolm J. Sheardown[†] and Christian Thomsen[‡]

Departments of Medicinal Chemistry, [†]Neuropharmacology and [‡]Receptor Neurochemistry,
Drug Discovery, Novo Nordisk Pharmaceuticals Division,
Novo Nordisk Park, DK 2760 Måløv, Denmark.

(Received in USA 11 June 1993; accepted 7 September 1993)

Abstract: The synthesis and SAR of a series of novel derivatives of *N*-aminoadenosine is described, along with their *in vitro* effects in biochemical assays. The rat brain A₁ adenosine receptor binding of these compounds is very dependent upon the purine 2-substituent. The novel agonist, 2-chloro-*N*-[4-(phenylthio)-1-piperidinyl]adenosine, exhibits a K_i value for A₁ receptor binding of <1 nM.

Adenosine 1 is a purine nucleoside with a wide variety of regulatory functions¹ and physiological effects². In recent years, the central nervous system (CNS) effects of this purine have received greater prominence³. It is now clear that in the mammalian CNS, adenosine acts as a neuromodulator⁴, and has been implicated both as an endogenous anticonvulsant agent⁵ and a neuroprotectant⁶. Agonists at adenosine receptors have therefore been shown to possess anticonvulsant⁷, antiischaemic⁸ and antinociceptive⁹ effects in laboratory animals, and have therefore been investigated for their therapeutic potential¹⁰ in treatment of human CNS disorders.



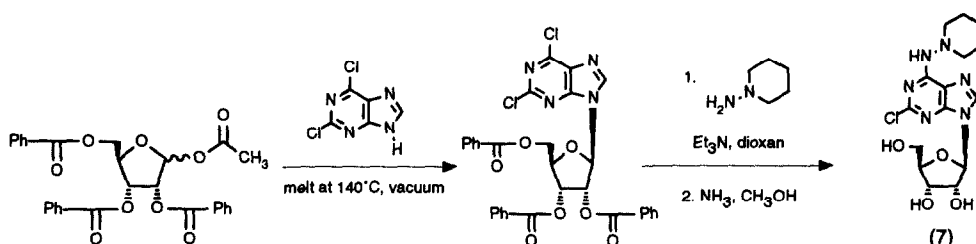
The challenge facing groups involved in designing novel A₁ receptor agonists with a view to the discovery of new drugs is to provide patentable compounds within an extensively investigated field, providing therapeutic benefit without limiting side effects. Despite over three decades of research within academia and the healthcare industry, the only adenosine receptor agonist to have been successfully registered so far as a drug for human use is adenosine itself, for treatment of arrhythmias, although several other marketed drugs have purinergic components to their action.

Some potent A₁ receptor agonists currently available include *N*-cyclopentyladenosine (CPA) 2, as well as its cyclohexyl analogue CHA¹⁰ and (2*S*)-*N*-(2-endo-norbornyl)adenosine [*S*(-)-ENBA]¹¹.

2-Chloro-*N*-cyclopentyladenosine (CCPA) **3** is claimed to be one of the most selective at the rat brain A₁ receptor¹². *R*-Phenylisopropyladenosine (*R*-PIA) **4** is a classical A₁ agonist, from which a group at Rhone Poulenc Rorer have derived the 3-chloro-2-thienyl derivative RG 14718(-) **5**, which is apparently the most potent A₁ ligand described to date¹³.

As part of our search for novel adenosine receptor agonists, we discovered that there are very few adenosine derivatives where an N-N bond exists at the purine 6-position¹⁴. In this communication we describe the synthesis, receptor binding and functional effect of a novel series of nucleosides of general structure **6**, which can be considered to be aza isosteres of reference A₁ receptor agonists such as CCPA **3**. Tables I - IV illustrate the effects of these compounds in biochemical assays comprising of adenosine A₁ and A₂ receptor binding, inhibition of cAMP accumulation and inhibition of contractile force in isolated guinea pig atria.

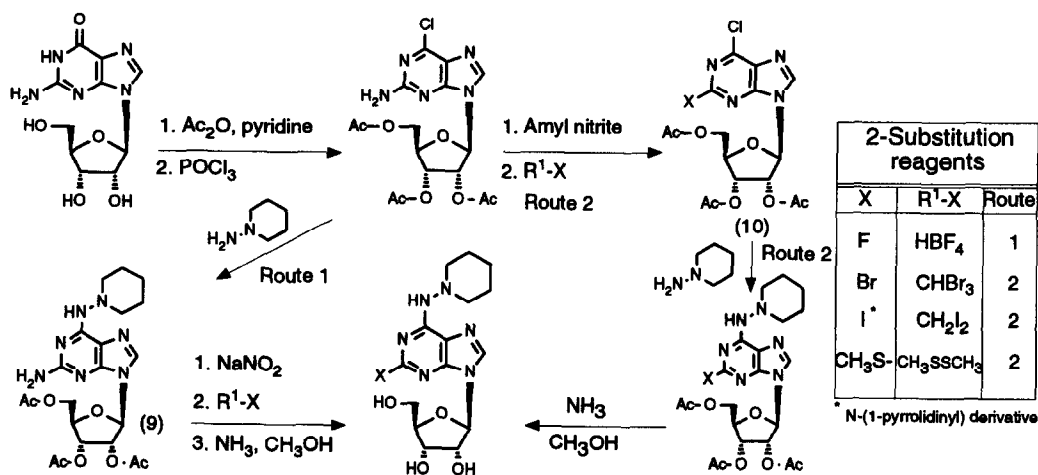
Scheme I: Synthesis of 2-chloro-*N*-(1-piperidinyl)adenosine.



Our initial target structure, 2-chloro-*N*-(1-piperidinyl)adenosine **7**, an aza analogue of 2-chloro-*N*-cyclohexyladenosine, was prepared as shown in Scheme 1. The above route^{15,16} was used to provide the 2-chloroadenosine derivatives in Tables I and III.

N-(1-piperidinyl)adenosine **8**, the deschloro analogue of **7**, was isolated by reaction of 1-amino-piperidine with 6-chloropurine riboside, and the surprising observation was made that this example showed no significant A₁ receptor binding (Table II). This was in marked contrast to **7**, which had a K_i value of 4nM for A₁ receptor binding (Table I), with high A₁/A₂ selectivity. This effect is not seen in *N*-alkyladenosine derivatives such as CCPA **3**, in which a 2-hydrogen substitution, giving CPA **2**, retains good binding affinity. The agonist **7** also showed potent effects in two functional assays indicative of A₁ agonist potency, inhibition of stimulated cAMP accumulation and inhibition of contractile force in isolated guinea pig heart (Table IV).

We therefore targeted a range of adenosine derivatives containing either a 1-piperidinyl or 1-pyrrolidinyl function at the 6-amino position, but with varying 2-substituents, in order to investigate what would be the optimal 2-substituent for A₁ agonist effect. A direct route from guanosine was used for most of these compounds¹⁷. Following ribosyl protection and chlorination at the purine 6-position, it was possible to substitute sequentially at either the 6-position to give **9**, or at the 2-position following diazotization of **10** (Scheme II).

Scheme II: Synthesis of 2-substituted adenosine derivatives from guanosine.

The rat brain adenosine receptor binding^{14,18} of this series of adenosine agonists was found to be very dependent upon the nature of the 2-substituent (Table II). Tables I and II show that whilst there is some freedom in the choice of the purine *N*-substituent, the requirements for purine 2-substituents

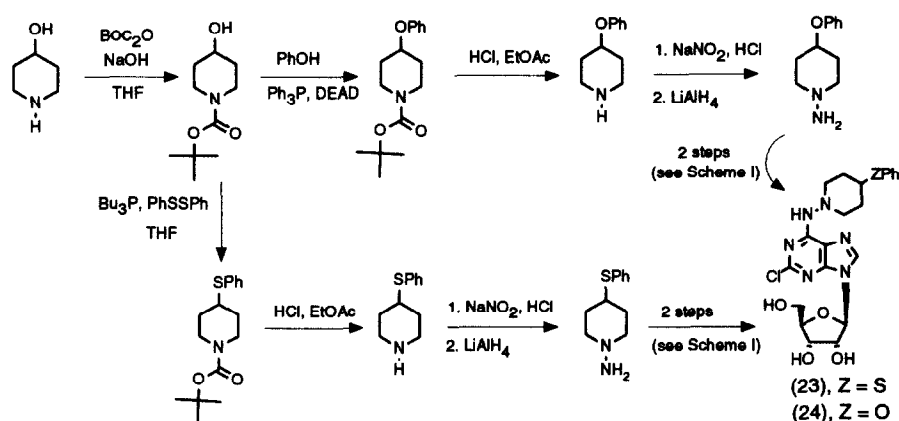
Table I: Adenosine Receptor binding and log P values¹⁹ of 2-chloroadenosine derivatives and Table II: Other 2-substituted adenosines.

Compound number	X	Adenosine N-substituent	Receptor binding		LogP
			A1	A2	
7	Cl	-N(CH ₂ OCH ₃)	4.2	1485	0.79
11	Cl	(S)-N(CH ₂ OCH ₃)	9.9	564	0.32
12	Cl	-N(CH ₃)	4.0	33	1.38
13	Cl	(R)-N(CH ₂ OCH ₃)	10.9	459	0.32
14	Cl	-N(CH ₃)	9.3	1130	1.42
15	Cl	-N(CH ₃)	13.3	1040	0.11
16	Cl	-N(CH ₃)	184	>30000	0.11
3	Cl	-N(CH ₃)	1.5	667	2.07

Compound number	X	Adenosine N-substituent	Receptor binding		LogP
			A1	A2	
17	F	-N(CH ₃)	3.6	1050	0.35
18	Br	-N(CH ₃)	9.3	1520	0.89
19	NH ₂	-N(CH ₃)	30	3320	-0.55
20	CH ₃ S	-N(CH ₃)	390	2825	0.89
21	n-PrO	-N(CH ₃)	400	2700	-
22	I	-N(CH ₃)	13000	57000	1.42
8	H	-N(CH ₃)	>3000	>10000	1.28
2	H	-N(CH ₃)	1.2	321	0.72

are more restrictive. In this series with cyclic hydrazines as purine 6-substituents, we found that electronegative halogens (F, Cl, Br) at the 2-position gave the strongest A_1 receptor binding. The receptor binding and agonist activity (Table IV) of this class of adenosine agonists could generally be increased further by substituting the *N*-(1-piperidinyl) group in the purine 6-position. We found that using the synthetic route illustrated in Scheme I, but with various phenyl-, phenoxy-, phenylthio- or phenylsulphonyl-substituted 1-aminopiperidines (Scheme III), some highly potent A_1 receptor agonists were obtained (Table III). The method used involved *N*-protection of 4-piperidinol, followed by either aryl ether²⁰ or thioether²¹ formation. The *N*-amination procedure utilized was the classical procedure involving nitrosation followed by reduction to a hydrazine²².











Scheme III. Synthesis of 2-chloro-*N*-[4-(phenylthio)-1-piperidinyl]adenosine and 2-chloro-*N*-[4-(phenoxy)-1-piperidinyl]adenosine.



Several conceptual steps were required to attain compound **23**, 2-chloro-*N*-[4-(phenylthio)-1-piperidinyl]adenosine, which is approximately equipotent in adenosine A_1 receptor binding to the apparently most potent commercially available A_1 agonist *S*(-)-ENBA¹¹. Using **32** as a starting point, potency was increased by preparing the deaza example **27**. Further potency increase was seen when a phenyl ether oxygen was introduced to give **24**, which continued in thioether **23**. Thus, the end result, introduction of a phenyl-containing substituent in the parent structure **7** in a spatial orientation which may give some overlap with the phenyl ring in e.g. *R*-PIA **4** gave a significant increase in the *in vitro* potency of this series of compounds.

For selected compounds, the effects in biochemical assays measuring inhibition of stimulated cAMP accumulation in smooth muscle cells²³ and lowering of the force of contraction in the isolated guinea pig atria²⁴ were determined (Table IV). These assays are known to give a good indication of adenosine A_1 functional effect in these two tissues. All the compounds tested in the cAMP assay displayed full agonist effect when compared to the reference A_1 agonist *R*-PIA **4**. The *in vivo* CNS characterization of these examples will be described elsewhere.

Table III: Adenosine Receptor binding and log P values of substituted 2-chloro-*N*-(1-piperidiny)adenosine derivatives and Table IV: Effects of selected compounds in functional assays.

Compound number	X	Adenosine N-substituent	Receptor binding A1 K _i , nM	A2	LogP
23	Cl		0.9	470	2.62
24	Cl		1.4	1100	2.05
25	Cl		6.1	638	2.28
26	Cl		15	1733	1.88
27	Cl		11	1200	2.36
28	Cl		11	720	-0.82
29	Cl		13	1380	2.89
30	Cl		17	220	-
31	Cl		34	17	2.25
32	Cl		55	5571	1.42

Compound number	Inhibition of stim. cAMP accum. IC ₅₀ (nM)	GP atria -lowering of contractile force IC ₅₀ (μM)
7	2.1	0.58
17	3.8	1.29
23	0.3	0.64
24	0.03	0.19
25	1.1	2.9
26	4.8	4.5
27	2.5	0.32
28	4.6	0.47
30	0.7	0.32
CPA	0.6	0.42
R-PIA	0.4	0.44
S-PIA	27	8.2

*mixture of diastereoisomers

In conclusion, we have described the synthesis, preliminary SAR and biochemical effects of a range of novel substituted *N*-aminoadenosine derivatives, some with very potent A₁ receptor binding and functional effect. These compounds are very unusual amongst adenosine receptor agonists in the apparent necessity of an electronegative halogen in the 2-position of the purine moiety for receptor binding; substitution with a hydrogen in the 2-position provided only very weakly active examples.

Acknowledgment: We wish to acknowledge the skilled contribution of Paw Bloch, Claus Jensen and Henrik Stephensen for the synthesis of novel nucleosides and Lisbeth Hansen, Iben Bredmose, Anne Meincke and Lone Igel for biochemical studies as well as Helle Kristensen and Lene Wendel for log P determinations. Furthermore, one of us (L.J.S.K.) wishes to thank Drs. Brian Evans and Frank Ellis of Glaxo Group Research, Ware, U.K. for helpful discussions.

References and Notes

1. *Adenosine and Adenine Nucleotides as Regulators of Cellular Function*; J.W. Phillis, Ed., CRC Press, Boca Raton, Fla., 1991.
2. Daval, J-L.; Nehlig, A.; Nicolas, F. Physiological and Pharmacological Properties of Adenosine: Therapeutic Implications. *Life Sci.*, **1991**, *49*, 1435-1453.
3. *Adenosine in the Nervous System*, T.W. Stone, Ed., Academic Press Ltd., London, 1991.
4. Williams, M.; Adenosine - a selective neuromodulator in the mammalian CNS? *Trends Neurosci.*, **1984**, 164-168.
5. Dragunow, M.; Goddard, G.V.; Lavery, R. Is Adenosine an Endogenous Anticonvulsant? *Epilepsia*, **1985**, *26*, 480-487.
6. Rudolph, K.A.; Schubert, P.; Parkinson, F.E.; Fredholm, B.B. Neuroprotective role of adenosine in cerebral ischaemia. *Trends Pharmacol. Sci.*, **1992**, *11*, 439-445.

7. Dunwiddie, T.V.; Worth, T. Sedative and anticonvulsant effects of adenosine analogs in mouse and rat. *J. Pharm. Exp. Ther.*, **1982**, *220*, 70-76; Klitgaard, H.; Knutsen, L.J.S.; Thomsen, C. Contrasting effects of adenosine A₁ and A₂ receptor ligands in different chemoconvulsive rodent models. *Eur. J. Pharmacol.*, **1993**, in press.
8. Marangos, P.J.; von Lubitz, D.; Daval, J-L; Deckert, J. Adenosine: its relevance to the treatment of brain ischemia and trauma. *Current and Future Trends in Anticonvulsant, Anxiety, and Stroke Therapy*; Wiley-Liss Inc., 1990; pp. 331-349.
9. Karlsten, R.; Post, C.; Hide, I.; Daly, J.W. The antinociceptive effect of intrathecally administered adenosine analogs in mice correlates with the affinity for the A₁-adenosine receptor. *Neurosci. Lett.*, **1991**, *121*, 267-270.
10. Jacobsen, K.A.; van Galen, P.J.M.; Williams, M. Adenosine Receptors: Pharmacology, Structure-Activity Relationships and Therapeutic Potential. *J. Med. Chem.*, **1992**, *35*, 407-422.
11. Trivedi, B.K.; Bridges, A.J.; Patt, W.C.; Priebe, S.R.; Bruns, R.F. N⁶-Bicycloalkyladenosines with Unusually High Potency and Selectivity for the Adenosine A₁ Receptor. *J. Med. Chem.*, **1989**, *32*, 8-11.
12. Lohse, M.J.; Klotz, K.N.; Schwabe, U.; Cristalli, G.; Vittori, S.; Grifantini, M. 2-Chloro-N⁶-cyclopentyladenosine: a highly selective agonists at A₁ adenosine receptors. *Naunyn-Schmiedeberg's Arch. Pharmacol.*, **1988**, *337*, 687-689.
13. Fink, C.A.; Spada, A.P.; Colussi, D.; Rivera, L.; Merkel, L. Synthesis of a potent A₁ Adenosine Receptor Agonist: N⁶-[1-R-[3-Chloro-2-thienyl)methyl]propyl]adenosine, RG 14718(-). *Nucleosides Nucleotides*, **1992**, *5*, 1077-1088.
14. a) Giner-Sorolla, A.; O'Bryant, S.A.; Nanos, M.R.; Dollinger, M.R.; Bendich, A.; Burchenal J.H. The Synthesis and Biological Properties of Hydroxylaminopurines and Related Derivatives. *J. Med. Chem.*, **1968**, *11*, 521-523. b) Montgomery J.A.; Hewson, K. 2-Fluoropurine Ribonucleosides. *J. Med. Chem.*, **1970**, *13*, 427-430. c) Kikugawa, K.; Iizuka, K.; Higuchi, Y.; Hirayama, H.; Ichino, M.; Platelet Aggregation Inhibitors. 2. Inhibition of Platelet Aggregation by 5'-, 2-, 6-, and 8-Substituted Adenosines. *J. Med. Chem.*, **1972**, *15*, 387-390. d) Kusachi, S.; Thompson, R.D.; Bugni, W.J.; Yamada, N. Dog Coronary Artery Adenosine Receptor: Structure of the N⁶-Alkyl Subregion. *J. Med. Chem.*, **1985**, *28*, 1636-1643. d) Saneyoshi, M.; Terashima, K. Synthetic Nucleosides and Nucleotides. VII. A Direct Replacement of 6-Thiol Group of 6-Thioinosine and 6-Thioguanosine with Hydrazine Hydrate. *Chem. Pharm. Bull.*, **1969**, *17*, 2373-2376. e) Nair, V.; Fasbender, A.J. C-2 Functionalized N⁶-cyclosubstituted Adenosines: Highly Selective Agonists for the Adenosine A₁ Receptor. *Tetrahedron*, **1993**, *49*, 2169-2184.
15. Knutsen, L.J.S.; Lau, J. Novel 2,6-Disubstituted Purine Derivatives. WO 93/08026 (29 April 1993).
16. Imai, K.; Nohara, A.; Honjo, M. Synthesis of Purine Nucleosides using Iodine as Catalyst. *Chem. Pharm. Bull.*, **1966**, *14*, 1377-1381. It was found that iodine was unnecessary in the first reaction in Scheme I.
17. Nair, V.; Young, D.A. A New Synthesis of Isoguanosine, *J. Org. Chem.*, **1985**, *50*, 406-408.
18. Williams, M.; Jacobsen, K.A. Radioligand Binding Assays for Adenosine Receptors. *Adenosine and Adenosine Receptors*; Williams, M., Ed.; The Humana Press Inc.: Clifton, N.J., 1990; pp 17-55. Binding with [³H]-CGS 21680 as radioligand may be termed A_{2b} receptor binding.
19. Log P values were determined either by the octanol/pH 7.4 buffer method or by HPLC correlation.
20. Mitsunobu, O. The Use of Diethylazodicarboxylate and Triphenylphosphine in Synthesis and Transformation of Natural Products. *Synthesis*, **1981**, 1-28.
21. Kotsuki, H.; Matsumoto, K.; Nishizawa, H. High Pressure-Promoted Transformations of Alcohols into the Corresponding Phenylsulphides with Bu₃P-PhSSPh. *Tetrahedron Lett.*, **1991**, *32*, 4155-4158.
22. Overberger, C.G.; Herin, L.P. Azo Compounds, XXXVIII. The Mercuric Oxide Oxidation of 1-Amino-2-phenylpiperidine. *J. Org. Chem.*, **1962**, *27*, 417-422.
23. Gerwins, P.; Norstedt, C.; Fredholm, B.B.; Characterization of Adenosine A₁ receptors in intact DDT₁ MF2 smooth muscle cells. *Mol Pharmacol.*, **1990**, *38*, 660-666.
24. For a method description, see Sauerberg, P.; Olesen, P.H.; Nielsen, S.; Treppendahl, S.; Sheardown, M.J.; Honoré, T.H.; Mitch, C.H.; Ward, J.S.; Pike, A.J.; Bymaster, F.; Sawyer, B.D.; Shannon, H.E. Novel Functional M₁ Selective Agonists. Synthesis and Structure-Activity Relationships of 3-(1,2,5-Thiadiazolyl)-1,2,5,6-tetrahydro-1-methylpyridines. *J. Med. Chem.*, **1992**, *35*, 2274-2283.